

Seed Germination of a Halophytic Grass Aeluropus lagopoides

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Received: 18 July 2000 Returned for revision: 25 September 2000 Accepted: 2 November 2000 Published electronically: 26 January 2001

Aeluropus lagopoides (Linn.) Trin. Ex Thw. (Poaceae) is a perennial grass distributed from coastal Sindh and Balochistan to saline flats of Punjab, Pakistan. Seeds collected from an inland population of A. lagopoides located on the University of Karachi campus were germinated under various levels of salinity (0, 100, 200, 300, 400 and 500 mM NaCl) and temperature regimes (10/20, 15/25, 20/30 and 25/35 °C) in a 12 h dark/12 h light photoperiod. Highest germination was obtained under non-saline conditions, and an increase in NaCl concentration progressively inhibited germination. Inhibition of germination was greater at cooler temperatures (10/20 °C) when no seed germinated above a concentration of 300 mM NaCl. The germination response at moderate temperatures (20/30 °C) was optimal, with 30 % of seeds germinating in 500 mM NaCl. The rate of germination decreased as salinity increased. Germination rate was highest at 20/30 °C and lowest at 10/20 °C. Seeds were transferred from salt solutions to distilled water after 20 d and those from high salinities recovered quickly at warmer temperatures with an optimal response at 20/30 °C.

Key words: Aeluropus lagopoides, germination, halophyte, Karachi, salinity, temperature.

INTRODUCTION

Halophytic grasses found on the Arabian Sea coast are relatively intolerant of high NaCl concentrations during germination. Halopyrum mucronatum and Sporobolus arabicus were found to germinate in up to 200 mm NaCl (Noor and Khan, 1995; Khan and Ungar, 2001) while Urochondra setulosa seeds germinated at 500 mm NaCl (Gulzar et al., 2001). Perennial halophytic grasses from other regions showed restricted germination at NaCl salinities approaching half strength seawater: Puccinellia nuttalliana at 344 mm (Macke and Ungar, 1971); Diplachne fusca at 400 mm (Morgan and Myers, 1989); Hordeum vulgare at 344 mm (Badger and Ungar, 1989); Briza maxima at 310 mm (Lombardi et al., 1998) and Typha latifolia at 310 mm (Lombardi et al., 1997), while some grasses like Spartina alterniflora showed some germination in 1027 mm NaCl (Mooring et al., 1971).

Salinity and temperature interact in their control of seed germination (Khan and Ungar, 1999), with the greatest inhibition due to salinity usually found at the minimum or maximum limits of tolerance to temperature (Badger and Ungar, 1989). The dependence of the response to salinity on temperature may be due to an initial inhibition of germination at higher salt concentrations at some temperatures rather than others (De Villiers *et al.*, 1994; Khan and Ungar, 1997, 1998, 1999; Khan and Gul, 1998), while alternating day and night temperatures have been shown

to promote germination in a number of halophytes (Okusanya, 1977; Ungar, 1995; Khan and Ungar, 1997, 1998, 1999; Gul and Weber, 1999).

Seeds of halophytes have been found to germinate even after prolonged exposure to hyper-saline conditions (Macke and Ungar, 1971; Woodell, 1985; Keiffer and Ungar, 1995, 1997). However, they differ in their capacity to recover from exposure to salinity (Keiffer and Ungar, 1995), a variation in recovery that could be due to differences in the temperature regimes to which seeds are exposed (Khan and Ungar, 1996, 1997, 1998, 1999). Gulzar *et al.* (2001) found that seeds of *U. setulosa* had 85% recovery when pretreated with 500 mm NaCl for 20 d at moderate temperatures (20/30 °C). Macke and Ungar (1971) found 87% recovery in distilled water as compared to 5·2% germination after 45 d of exposure to 344 mm NaCl in seeds of *Puccinellia nuttalliana*.

Aeluropus lagopoides (Linn.) Trin. Ex Thw. (Poaceae) is a salt-secreting rhizomatous perennial grass ranging in distribution from Northern Africa (Morocco to Somalia), Sicily and Cyprus, through the Middle East to Central Asia, Pakistan and India (Cope, 1982). The plant propagates vegetatively by rhizome growth after monsoon rains and it also produces numerous flowers and seeds from April to October (Gulzar and Khan, unpubl. res.). It is often found in association with Cressa cretica in inland communities and with Cyperus arenarius, Cressa cretica and Halopyrum mucronatum in coastal communities in the backwaters of the Manora creek, Pakistan. Aeluropus lagopoides is used throughout the year as a forage plant in parts of India. Its foliage shows little variation in salt content despite a three-fold increase in soil salinity in

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summer (Joshi and Bhoite, 1988); this is possibly due to salt secretion through leaf glands. The low salt content of *A. lagopoides* shoots is advantageous in its use as a fodder crop. Currently, little information is available regarding the relative salt tolerance of halophytic grasses from subtropical regions of the world. The primary objective of this investigation was to determine the response of *A. lagopoides* to saline conditions, to ascertain if *A. lagopoides* could germinate at NaCl concentrations approaching seawater (500 mm NaCl) and how this response might be governed by prevailing temperature conditions.

MATERIALS AND METHODS

Spikelets of Aeluropus lagopoides were harvested in winter 1997 from a population on the Karachi University campus. Seeds were separated from each inflorescence, cleaned, and dry-stored in a refrigerator at 4 °C after surface sterilization with 0.85 % clorox for 1 min. Germination experiments were initiated in September 1998. Six salinity concentrations (0, 100, 200, 300, 400 and 500 mm NaCl) were used, based on a preliminary test of salt tolerance. Germination was tested in a programmed incubator (Percival, Boone, USA) at (dark:light) 10/20, 15/25, 20/30 and 25/35 °C temperature regimes with a 12 h photoperiod (25 µmol m⁻² s⁻¹, 400–700 nm Sylvania cool white fluorescent lamps). In the inland community where A. lagopoides germinates, salinity is usually around 300 mm, decreasing to about 100 mm depending on the extent and duration of rainfall. Average day and night-time temperatures during the monsoon period are around 30 and 20 °C, respectively. Seeds were germinated in two folds of Whatman No. 1 filter paper placed in 2.5 cm × 18 cm glass test tubes with 5 ml of test solution. The tubes were sealed using parafilm. A seed was considered to have germinated at the emergence of the radicle. Germination was noted on alternate days. After 20 d, all non-germinated seeds were placed in distilled water at their initial temperature regimes for another 20 d. Rate of germination was calculated with a modified Timson's germination velocity index: $\sum G/t$, where G is the percentage of seed germinated after 2 d intervals, and t is the total time of germination (Khan and Ungar, 1998). Rate of recovery of germination was calculated using the relation: $\frac{(a-b)}{c}$ b)] 100, where a is the total number of seeds germinated after being transferred to distilled water, b is the total number of seeds germinated in saline solution and c is the total number of seeds. Germination data (20 d) and recovery germination data (20 d) were arcsine transformed before statistical analysis to ensure homogeneity of variance. Data were analysed using SPSS, version 9.0 (SPSS, 1999). The effect of salinity and temperature on germination, rate of germination and recovery of germination were examined using two-way ANOVA and regression analysis.

RESULTS

Two-way ANOVA indicated that germination of *A. lagopoides* seeds was significantly affected by temperature, salinity and their interaction (Table 1). Seed germina-

TABLE 1. Effect of salinity (S), temperature (T) and their interaction on generation of A. lagopoides seeds

Source of variation	S	T	SXT
Germination	131.7	129-1	3.4
Rate of germination	141.2	156-4	8.0
Recovery germination	39-3	82.6	14.3

Data are F values (two-way ANOVA) significant at P < 0.001.

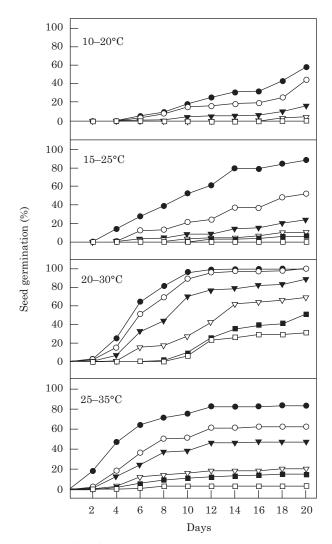


FIG. 1. The effect of temperature regime and salinity on the percentage germination over time of *Aeluropus lagopoides* seeds. NaCl concentration (mm): ●, 0; ○, 100; ▼, 200; ■, 400; □, 500.

tion was highest in distilled water in the 20/30 °C temperature regime (Fig. 1). Thirty percent of the seeds germinated in 500 mM NaCl at 20/30 °C while no seed germinated at 10/20 °C (Figs 1 and 2). Germination was affected by temperature and was reduced from 100 % at 20/30 °C to only 60 % at 10/20 °C in the non-saline control. Seeds showed higher germination in all treatments at 15/25 °C and 20/30 °C, with 100 % germination in the

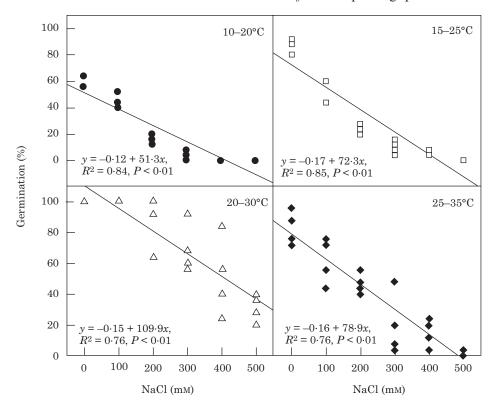


FIG. 2. Regression plots for mean final germination percentage of Aeluropus lagopoides seeds in different salinity and temperature regimes.

non-saline control; however, a further increase in temperature $(25/35 \, ^{\circ}\text{C})$ inhibited germination.

The rate of germination (calculated using a modified Timson's index) was highest at 20/30 °C and lowest at 10/20 °C (Fig. 3). Addition of NaCl to the medium adversely affected the germination rate at all temperatures, with a less marked response at 10/20 °C and 15/25 °C where non-saline control values were lower (Fig. 3). Twoway ANOVA indicated a significant effect of salinity and temperature on rate of germination (Table 1).

When seeds were transferred to distilled water after 20 d of salinity treatment, the recovery of germination percentages increased with an increase in pre-transfer salinity treatments except for $10/20~^{\circ}\text{C}$ (Fig. 4). Seeds subjected to high temperature had higher recovery percentages (Fig. 4). Seed germination recovered completely in distilled water at higher temperatures and at all salinity treatments, but recovery was lower (25 %) at $10/20~^{\circ}\text{C}$. A two-way ANOVA indicated significant (P < 0.001) effects of both salinity and temperature on total germination (Table 1).

DISCUSSION

Halophyte seeds germinate best under non-saline conditions (Ungar, 1995). Salinity inhibits germination of halophyte seeds in one of two ways: (1) preventing germination without loss of viability at higher salinities; and (2) delaying germination of seeds at salinities that cause some stress to seeds but do not prevent germination. The threshold salinity for a significant reduction in germination varies between grass species from 100–500 mm NaCl

(Lombardi et al., 1998; Gulzar et al., 2001; Khan and Ungar, 2001). The germination tolerance of plant species to salinity under laboratory conditions does not necessarily correlate with their response to salinity under field conditions, and may be many times lower. Seeds usually germinate after leaching of salt due to the monsoon rainfall in the vicinity of Karachi (Khan and Gul, 1998). Mahmood and Malik (1996) found that the grasses Sporobolus arabicus, Cynodon dactylon, Polypogon monspeliensis and Desmostachya bipinnata showed a decrease in germination with an increase in salinity from 3-20 dS m⁻¹. Only D. bipinnata exhibited salt stimulation at low salinity (5 dS m⁻¹ or 50·16 mm NaCl) and had the highest (40 %) germination at 20 dS m⁻¹ (206 mm NaCl) with no seeds germinating at higher salinities. Hyder and Yasmin (1972) found that the germination of Sporobolus airoides seeds was inhibited by a specific ion effect. They exhibited 82 % germination in distilled water and 2 % in 325 mm NaCl. Gulzar et al. (2001) reported that seeds of U. setulosa could germinate in up to 500 mm NaCl while those of H. mucronatum failed to germinate at 300 mm NaCl (Noor and Khan, 1995). Aeluropus lagopoides seeds showed no dormancy under non-saline conditions. An increase in salinity inhibited germination; however, about 30 % of the seeds germinated at 500 mm NaCl. These results indicate that A. lagopoides is a highly salt tolerant grass in comparison to other grasses found at inland and coastal sites near Karachi.

Sensitivity to periodic temperature and salinity fluctuations constitutes an important system which enables plants to respond to daily variations in the soil surface conditions.

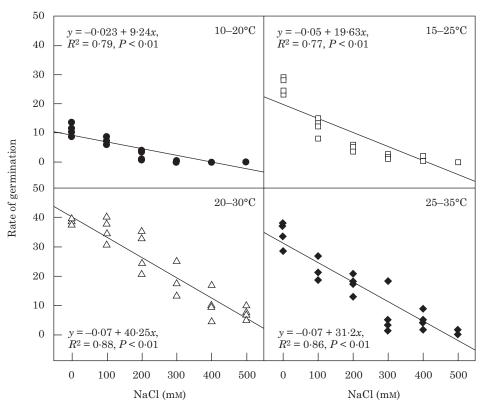


FIG. 3. Regression plots for the rate of germination (maximum = 50) of Aehropus lagopoides seeds in different salinity and temperature regimes.

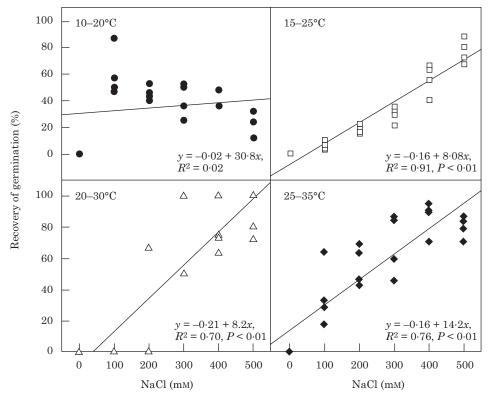


FIG. 4. Regression plots for percentage recovery germination of Aeluropus lagopoides seeds in different salinity and temperature regimes.

Temperature shifts may affect a number of processes determining the germinability of seeds, including membrane permeability, activity of membrane-bound proteins, and cytosol enzymes (Bewley and Black, 1994). The response of halophytic seeds to alternating temperatures and soil salinity levels is of ecological significance. Germination of halophyte seeds in subtropical coastal and inland salt marshes usually occurs after monsoon rains when there is a reduction in temperature and soil salinity (Khan and Ungar, 1996, 1997, 1998, 1999; Khan and Gul, 1998). Salinity and temperature interact in their control of seed germination and the greatest inhibition is usually found at the maximum and minimum limits of tolerance of these two environmental variables, as reported in Hordeum jubatum (Badger and Ungar, 1989), Crambe abyssinica (Fowler, 1991), Puccinellia ciliata (Myers and Couper, 1989), Halopyrum mucronatum (Noor and Khan, 1995), and Urochondra setulosa (Gulzar et al., 2001). The effect of salinity on germination varies considerably with temperature (Fowler, 1986; Badger and Ungar, 1989; Morgan and Myers, 1989; Myers and Morgan, 1989; Romo, 1990; Gutterman, 1993; Noor and Khan, 1995; Gulzar et al., 2001). Cluff et al. (1983) reported that a temperature regime of 10/40 °C (16/8 h) is optimal for seed germination of Distichlis spicata, whereas -5 and 50 °C were the lower and upper threshold temperatures for seed germination. Only 1 % germination was observed in −15 bars NaCl concentration at the optimum temperature of 10/40 °C. Aeluropus lagopoides showed similar results with 31 % germination at the optimal temperature under the highest salinity treatment (500 mm NaCl), but an increase or decrease in temperature regime significantly inhibited germination at all salinities. This is in contrast with Arthrocnemum macrostachyum (Khan and Gul, 1998) and Suaeda fruticosa (Khan and Ungar, 1998), where temperature variations were not significant at lower salinities.

The response of seeds of *A. lagopoides* transferred to distilled water after 20 d at various salinities varied depending on the temperature regime. Seeds exposed to higher salinity recovered quickly at warmer temperatures. Under hyper-saline conditions, seed survival rather than germinability may be an appropriate criterion for success, since recovery germination does occur in seeds of *A. lagopoides* and other halophytes when hyper-saline conditions are alleviated (Ungar, 1995; Khan and Ungar, 1996, 1997, 1998, 1999). Recovery germination responses were also dependent on temperature, ranging from 29 % recovery at 10/20 °C to 85 % at 20/30 °C.

Aeluropus lagopoides is a perennial grass with an extensive rhizome system that can tolerate high salinities (Bodla et al., 1995). Coastal populations are exposed to seasonal tidal inundation and a highly saline shallow water table. Inland populations grow in highly saline wet soils. Aeluropus lagopoides propagates primarily through rhizomes, although it produces a large number of seeds and maintains a transient seed bank (Khan and Gul, 1999). The present study demonstrates that seeds are not dormant and showed 100 % germination at the optimal temperature. Seeds of A. lagopoides should be considered among the most salt tolerant of local grasses found in saline soils in the

Karachi region. Thirty percent of seeds germinated at 500 mm NaCl, a salinity concentration approaching seawater (600 mm NaCl). These data showed that *A. lagopoides* could be recruited easily in highly saline inland and coastal conditions under warm temperatures. It seems that the proximate strategy of *A. lagopoides* is to use vegetative methods for recruitment of new individuals—a less costly way of recruitment in a highly unpredictable, harsh environment. Production of a large number of seeds is probably an ultimate strategy to introduce new genotypes into *A. lagopoides* populations at an appropriate time to maximize fitness. Further investigations are necessary to understand the ecophysiological strategies of plants for survival under natural environmental conditions.

ACKNOWLEDGEMENT

We thank the University of Karachi for providing the research grant.

LITERATURE CITED

- **Badger KS, Ungar IA. 1989.** The effects of salinity and temperature on the germination of the inland halophyte *Hordeum jubatum. Canadian Journal of Botany* **67**: 1420–1425.
- **Bewley JD, Black M. 1994.** Seeds: Physiology of development and germination. London: Plenum Press.
- Bodla MA, Choudhry MR, Shamsi SRA, Baig MS. 1995. Salt tolerance in some dominant grasses of Punjab. In: Khan MA, Ungar IA, eds. *Biology of salt tolerant plants*. Karachi, Pakistan: University of Karachi, 190–198.
- Cluff GJ, Evans RA, Young JA. 1983. Desert salt grass seed germination and seedbed ecology. *Journal of Range Management* 36: 419–422.
- Cope TA. 1982. Poaceae. In: Nasir E, Ali SI, eds. Flora of Pakistan. Karachi, Pakistan: University of Karachi.
- De Villiers AJ, Van Rooyen MW, Theron GK, van de Venter HA. 1994. Germination of three Namaqualand pioneer species as influenced by salinity, temperature and light. Seed Science and Technology 22: 427–433.
- **Fowler JL. 1991.** Interaction of salinity and temperature on the germination of *Crambe abyssinica*. *Agronomy Journal* **83**: 169–172.
- Fowler NL. 1986. Microsite requirements for germination and establishment of three grass species. *American Midland Naturalist* 115: 131–145.
- **Gul B, Weber DJ. 1999.** Effect of salinity, light and temperature on germination in *Allenrolfea occidentalis. Canadian Journal of Botany* 77: 240–246.
- Gulzar S, Khan MA, Ungar IA. 2001. Effect of salinity and temperature on the germination of *Urochondra setulosa*. Seed Science and Technology (in press).
- Gutterman Y. 1993. Seed germination of desert plants. Berlin: Springer-Verlag.
- Hyder SZ, Yasmin S. 1972. Salt tolerance and cation interaction in alkali sacaton at germination. *Journal of Range Management* 25: 390-392.
- **Joshi AJ, Bhoite AS. 1988.** Fluctuations of mineral ions in saline soils and halophytic grass *Aeluropus lagopoides* L. *Annals of Arid Zone* **27**: 191–196.
- Keiffer CH, Ungar IA. 1995. Germination responses of halophyte seeds exposed to prolonged hypersaline conditions. In: Khan MA, Ungar IA, eds. Biology of salt tolerant plants. Karachi, Pakistan: University of Karachi, 43–50.
- **Keiffer CH, Ungar IA. 1997.** The effect of extended exposure to hypersaline conditions on the germination of five inland halophyte species. *American Journal of Botany* **84**: 104–111.

- **Khan MA, Gul B. 1998.** High salt tolerance in the germinating dimorphic seeds of *Arthrocnemum indicum*. *International Journal of Plant Sciences* **159**: 826–832.
- Khan MA, Gul B. 1999. Seed bank strategies of coastal populations at Arabian sea coast. In: McArthur ED, Ostler WK, Wambolt CL, eds. Proceedings of the symposium on Shrubland Ecotones. Ogden: USDA Forest Service, Rocky Mountain Research Station, 227–230.
- Khan MA, Ungar IA. 1996. Influence of salinity and temperature on the germination of *Haloxylon recurvum*. Annals of Botany 78: 547-551
- Khan MA, Ungar IA. 1997. Effect of temperature regime on recovery of seed germination of halophytes from saline conditions. *American Journal of Botany* 84: 279–283.
- Khan MA, Ungar IA. 1998. Germination of salt tolerant shrub Suaeda fruticosa from Pakistan: Salinity and temperature responses. Seed Science and Technology 26: 657–667.
- **Khan MA, Ungar IA. 1999.** Effect of salinity on the seed germination of *Triglochin maritima* under various temperature regimes. *Great Basin Naturalist* **59**: 144–150.
- **Khan MA, Ungar IA. 2001.** Role of dormancy regulating chemicals in release of innate and salinity-induced dormancy in *Sporobolus arabicus* Boiss. *Seed Science and Technology* (in press).
- **Lombardi T, Fochetti T, Andrea B, Onnis A. 1997.** Germination requirements in a population of *Typha latifolia. Aquatic Botany* **56**: 1–10.
- Lombardi T, Fochetti T, Onnis A. 1998. Germination of *Briza maxima*L. seeds: effects of temperature, light, salinity and seed harvesting time. *Seed Science and Technology* 26: 463–470.
- Macke A, Ungar IA. 1971. The effect of salinity on germination and early growth of *Puccinellia nuttalliana*. Canadian Journal of Botany 49: 515–520.

- Mahmood K, Malik KA. 1996. Seed germination and salinity tolerance in plant species growing on saline wastelands. *Biologia Plantarum* 38: 309–315.
- Mooring MT, Cooper AW, Seneca ED. 1971. Seed germination response and evidence for height of ecophenes in *Spartina alterniflora* from North Carolina. *American Journal of Botany* 58: 48–56.
- Morgan WC, Myers BA. 1989. Germination of the salt-tolerant grass *Diplachne fusca*. I. Dormancy and temperature responses. *Australian Journal of Botany* 37: 225–237.
- Myers BA, Couper DI. 1989. Effects of temperature and salinity on the germination of *Puccinellia ciliata* (Bor.) cv. Menemen. *Australian Journal of Agricultural Research* 40: 561–571.
- Myers BA, Morgan WC. 1989. Germination of the salt-tolerant grass Diplachne fusca. II. Salinity responses. Australian Journal of Botany 37: 239–251.
- Noor M, Khan MA. 1995. Factors affecting germination of summer and winter seeds of *Halopyrum mucronatum* under salt stress. In: Khan MA, Ungar IA, eds. *Biology of salt tolerant plants*. Karachi, Pakistan: University of Karachi, 51–58.
- **Okusanya OT. 1977.** The effect of seawater and temperature on the germination behavior of *Crithmum maritimum. Physiologia Plantarum* **41**: 265–267.
- Romo JT. 1990. Stratification, freezing and drying effects on germination and seedling growth of *Altai wild rye. Journal of Range Management* 43: 167–171.
- SPSS. 1999. SPSS 9.0 for Windows Update. SPSS Inc. USA.
- Ungar IA. 1995. Seed germination and seed-bank ecology in halophytes. In: Kigel J, Galili G, eds. Seed development and seed germination. New York: Marcel Dekker, 599–628.
- **Woodell SRJ. 1985.** Salinity and seed germination patterns in coastal plants. *Vegetatio* **61**: 223–230.